SYNTHESIS OF OPTICALLY ACTIVE FORMS OF A-FACTOR, THE INDUCER OF STREPTOMYCIN BIOSYNTHESIS IN INACTIVE MUTANTS OF STREPTOMYCES GRISEUS[†]

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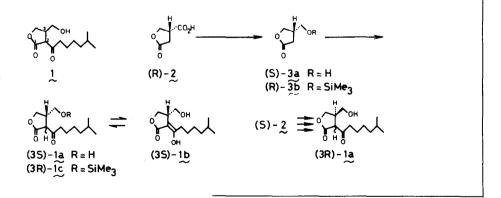
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Abstract—The absolute configuration at C-3 of A-factor, 2-(6-methylheptanoyl)-3-hydroxymethyl - 4 - butanolide 1a, was determined to be S by synthesizing both enantiomers of it from paraconic acid. (3S)-A-factor was 2.5 times more active than the (3R)-isomer as the inducer of streptomycin biosynthesis in inactive mutants of *Streptomyces* griseus.

Khokhlov et al. discovered A-factor as the inducer of the biosynthesis of streptomycin in inactive mutants of Streptomyces griseus.¹ It also induces the formation of spores in asporophological modifications of S. griseus.¹ In 1976, Russian workers proposed the gross structure of A-factor to be 1 by chemical and spectroscopic studies,² which was later confirmed by a syn-thesis of $(\pm)-1$.³ They also deduced the absolute stereochemistry of A-factor to be 2S, 3S as shown in 1a.^{2,4}[‡] This was done on the basis of the application of the Klyne lactone sector rule⁵ to this system 1. Since A-factor can exist either as a keto-form 1a or as an enol-form 1b, the applicability of the lactone sector rule is not unambiguous. We therefore decided to synthesize both enantiomers of A-factor from an optically active compound with known absolute stereochemistry in order to establish the configuration at C-3 unambiguously.⁶ For this purpose paraconic acid 2 seemed to be the most appropriate starting material, because the absolute configuration of (-)-2 was known to be R by its conversion to (S)-(-)-methylsuccinic acid.⁷

 (\pm) -Paraconic acid 2 was resolved carefully with (R)-(+)- α -phenethylamine to give a pure crystalline salt. This was dissolved in MeOH-H₂O and filtered through Amberlite IR-120 (H⁺-form) to give (R)-(~)-2, m.p. 57-58°, $[\alpha]_{D}^{23}$ - 59.6° (MeOH), (lit⁷ m.p. 48°, $[\alpha]_{D}^{30}$ + 49° (MeOH) for (S)-(+)-2), after two recrystallizations from CHCl₃. Reduction of (R)-(-)-paraconic acid 2 with BH₃ · Me₂S in THF afforded (S)-(-)-3-hydroxymethyl-4butanolide 3a. Unfortunately attempts to determine the optical purity of (S)-(-)-3a by analyzing it or its (S)-(-)-3a $(\alpha$ -methoxy- α -trifluoromethylphenyl-MTPA ester acetate) were unsuccessful. The hydroxy lactone 3a as well as A-factor 1a itself was highly susceptible to racemization in the presence of a base or an acid due to intramolecular transesterification. Careless work-up of 3a or 1a led to partial or complete racemization. To avoid any racemization, the OH group of (S)-3a was protected as Me₃Si ether (R)-(-)-3b by treatment with (Me₃Si)₂NH and Me₃SiCl in C₃H₃N. The lactone enolate generated by treating (R)-(-)-3b with 2.5 eq of LiNPr₂ⁱ in THF at - 78° was acylated with 6-methylheptanoyl chloride at



[†]Synthetic microbial chemistry—II. Part I, K. Mori and K. Sato, *Tetrahedron* **38**, 1221 (1982). The experimental part on the synthesis of (3R)-A-factor was taken from the B.Sc. thesis of K. Yamane (March 1982).

[‡]In their preliminary communication, Khokhlov *et al.* reported a 2S, 3R-stereochemistry for A-factor, while their formula in that paper showed a 3S-configuration.² They later revised this to the correct 2S, 3S-stereochemistry.⁴

In our preliminary communication⁶ we erroneously wrote (3S)-(+)-A-factor which should read (3S)-(-)-A-factor.

- 78° to give (3R)-1c. This was dissolved in EtOH-H₂O and the soln was heated under reflux to remove the Me₃Si protective group. Subsequent work-up followed by chromatographic purification afforded (3S)-(-)-Afactor 1a as a waxy solid, $[\alpha]_D^{23} - 13.1^\circ$ (CHCl₃); CD(MeOH) 283.5 nm ($\Delta \epsilon + 0.699$), 221 nm ($\Delta \epsilon + 0.420$).§ The natural A-factor was reported to show the following CD spectrum: CD(MeOH) 285 nm ($\epsilon + 0.349$), 225 nm ($\epsilon + 0.215$).² The comparison of these CD data proved the

stereochemical identity of (3S)-(-)-A-factor 1a with the natural product. This established beyond doubt the 3Sconfiguration of A-factor. Interestingly the CD data also suggested the higher optical purity of our synthetic Afactor than that of the natural one. Since the C-2 of (3S)-(-)-1a is readily enolizable via keto-enol tautomerization, the configuration at that position is uncertain. As suggested by Khokhlov, (2S, 3S)-1a seems to be thermodynamically more stable than (2R, 3S)-1a.^{2,4}

So as to synthesize the unnatural enantiomer (3R)-(+)-1a of A-factor, (\pm) -paraconic acid 2 was resolved with $(S)-(-)-\alpha$ -phenethylamine to give (S)-(+)-2, m.p. 60-63°, $[\alpha]_D^{21.5} + 61.7^\circ$ (MeOH). In a similar fashion to that described above for (3S)-(-)-1a, (S)-(+)-paraconic acid 2 yielded (3R)-(+)-A-factor 1a, $[\alpha]_D^{22} + 12.7^\circ$ (CHCl₃). Its CD spectrum was antipodal to that of (3S)-(-)-1a.

Bioassay of the enantiomers of A-factor was kindly carried out by Prof. T. Beppu and Mr. O. Hara of this Department. The least concentration effective as the inducer of streptomycin biosynthesis in inactive mutants of Streptomyces griseus was 2 ng/disc for (3S)-(-)-Afactor and 5 ng/disc for the (3R)-(+)-isomer. This means that the unnatural enantiomer also possesses weak biological activity (40% of the natural product).

EXPERIMENTAL

All b.ps were uncorrected. IR spectra were determined as film or as Nujol mull on a Jasco A-102 spectrometer. NMR spectra were recorded at 60 MHz with TMS as an internal standard on a Hitachi R-24A spectrometer unless otherwise stated. Optical rotations were measured on a Jasco DIP-140 automatic polarimeter.

Paraconic acid 2

(a) Racemate. To a stirred and heated soln of ethyl (\pm) paraconate⁷ (81 g) in 95% EtOH (170 ml) was added dropwise a soln of KOH (60 g) in H₂O (590 ml) during 1 h keeping the soln at reflux temp. After the addition the mixture was stirred and heated under reflux for 15 min. After cooling, the soln was slowly filtered through a column of Amberlite IR-120 (H⁺-form, 21.). The resin was washed with H₂O. The eluant (51.) was concentrated continuously with a flash evaporator. The residue was distilled to give a fraction with b.p. 170-180°/1.2 mm. This was mixed with CHCl₃ and left to stand in a refrigerator to give 47 g (70.6%) of crude (±)-2, m.p. 47-54°. Recrystallization of the crude product from CHCl₃ yielded prisms, m.p. 58-61.5° (lit⁷ m.p. 60-63°), $\begin{array}{l} \nu_{max} \sim 3200 \ (m), \ \sim 2600 \ (m), \ \sim 1735 \ (br. \ s), \ 1270 \ (m), \ 1230 \ (s), \\ 1190 \ (s), \ 1100 \ (w), \ 1030 \ (s), \ 1000 \ (m), \ 860 \ (m), \ 840 \ (m) \ cm^{-1}; \end{array}$ δ (CDCl₃) 2.79 (1H, d, J = 2 Hz), 2.92 (1H, s), 3.30–3.80 (1H, m), 4.55 (2H, d, J = 6 Hz), 10.52 (1H, s). This acid is hygroscopic. (Found: C, 45.74; H, 4.54. Calc. for C₅H₆O₄: C, 46.16; H, 4.65%.)

(b) (R)-(-)-Isomer. (R)-(+)- α -Phenethylamine (44 g) was added to a soln of (\pm) -2 (47 g) in MeOH (150 ml). Ether (50 ml) was added to the soln and the mixture was left to stand at room temp. The separated crystalline salt (52 g) was collected on a filter and recrystallized from MeOH-ether to give 29 g of the salt, $[\alpha]_D^{21}$ - 13.4° (c = 1.12, MeOH). This was recrystallized from MeOHether to give 18 g of the salt, $[\alpha]_D^{21} - 17.3^\circ$ (c = 1.19, MeOH). Further recrystallization from MeOH-ether gave 11 g of crystals, $[\alpha]_D^{21} - 20.6^\circ$ (c = 1.06, MeOH). This was again recrystallized from MeOH-ether to give 7 g (15.4%) of the (R)-acid-(R)-amine salt as needles, m.p. 150-152°, $[\alpha]_D^{21} - 25.0^\circ$ (c = 0.80, MeOH); $\nu_{max} \sim 2650$ (m), 2520 (w), 2200 (w), 1775 (s), 1625 (m), 1455 (m), 1395 (m), 1370 (m) 1170 (m), 1160 (m), 1020 (m), 770 (m), 700 (m) cm⁻¹. (Found: C, 61.98; H, 6.64; N, 5.57. Calc. for C₁₃H₁₇O₄N: C, 62.14; H, 6.82; N, 5.57%.) This salt (7 g) was dissolved in MeOH (20 ml)-H₂O (50 ml). The soln was filtered through Amberlite IR-120 (H⁺-form, 100 g). The resin was washed with H_2O (500 ml). The combined filtrate and washings were concentrated in vacuo with a flash evaporator. The residue

was dissolved in THF-C₆H₆ and concentrated in vacuo. This was repeated several times to remove H₂O. The residue was dissolved in THF-CHCh. The soln was filtered to remove solid impurities and the filtrate was concentrated in vacuo. The residue was recrystallized from CHCl₃ to give 2.5 g (69%) of (R)-2. Further recrystallization from CHCl₃ yielded 1.39g (39% from the salt) of (R)-2 as hygroscopic prisms, m.p. 57-58°, $[\alpha]_D^{23}$ - 59.6° (c = 0.614, MeOH). Because of its hygroscopic nature (R)-2 did not give correct analytical data. (Found: C, 45.03; H, 4.73. Calc. for C₅H₆O₄: C, 46.16; H, 4.65%.) The IR and NMR spectra were identical to those of the racemate.

(c) (S)-(+)-Isomer. (S)-(-)- α -Phenethylamine (59 g) was added to a soln of (\pm) -2 (62.7 g) in MeOH (170 ml). Ether was added to the soln and the mixture was left to stand overnight at room temp. The precipitated salt was recrystallized six times from i-PrOH to give 7 g (10.8%) of the (S)-acid-(S)-amine salt, m.p. 138–141°, $[\alpha]_D^{23} + 24.0^\circ$ (c = 1.032, MeOH). The IR spectrum was identical to that of the (R)-acid-(R)-amine salt. (Found: C, 62.36; H, 6.91; N, 5.52. Calc. for C13H17O4N: C, 62.14; H, 6.82; N, 5.57%.) This salt (13.11 g; $[\alpha]_d^{23} + 23.8^\circ$ (c = 1.016, MeOH)) was dissolved in MeOH (47 ml)-H2O (50 ml) and filtered through Amberlite IR-120 (H⁺-form, 200 g). The resin was washed with H₂O (11.). The combined filtrate and washings were concentrated in vacuo with a flash evaporator. The residue was worked up as described above for (R)-2 to give 3.55 g (52.3% from the salt) of (S)-2, m.p. 60-63°, $[\alpha]_{D}^{21.5}$ + 61.7° (c = 0.625, MeOH), as hygroscopic prisms. (Found: C, 45.56; H, 4.82. Calc. for C₅H₆O₄: C, 46.16; H, 4.65%.) The IR and NMR spectra were identical with those of the racemate.

3-Hydroxymethyl-4-butanolide 3a

(a) Racemate. BH3 · Me2S (1 ml) was added dropwise to a stirred and ice-cooled soln of (\pm) -2 (1.09 g) in dry THF (7.5 ml) under Ar at 0-5. The mixture was stirred for 3 h at 0-5°. Then MeOH (10 ml) was added dropwise with cooling to destroy BH₃. The soln was concentrated in vacuo. The residue was dissolved in MeOH (10 ml) and concentrated in vacuo. This was repeated once more. The residue was distilled to give 897 mg (92%) of (\pm) -3a, b.p. 160°/8 mm, n_D^{21} 1.4608; ν_{max} 3400 (br. s), 2930 (s), 2860 (s), 1760 (vs), 1475 (m), 1415 (m), 1380 (m), 1340 (m), 1290 (m), 1260 (m), 1180 (s), 1045 (s), 1020 (s), 970 (m), 940 (m), 895 (w), 835 (w) cm⁻¹; δ (CDCl₃) 2.3-2.9 (3H, m), 3.65 (2H, d, J = 6 Hz), 3.90 (1H, s), 4.1–4.6 (2H, m). (Found: C, 51.64; H, 7.36. Calc. for C₅H₈O₃: C, 51.72; H, 6.94%).

(b) (S)-(-)-Isomer. BH₃ · Me₂S (1.5 ml) was added dropwise to a stirred and ice-cooled soln of (R)-(-)-2 (1.3 g, $[\alpha]_D^{23}$ -59.6° (MeOH)) in dry THF (10 ml) under Ar at 0-5°. The mixture was stirred at 0-5° for 1 h and quenched with MeOH (5 ml). The soln was concentrated in vacuo below 30°. The residue was dissolved in MeOH (10 ml) and concentrated in vacuo. This was repeated again. The residue was chromatographed over SiO₂ (Mallinckrodt Silicar CC-7, 3 g) in n-hexane. Elution with CHCl₃ yielded 995 mg (89%) of (S)-3a, n_D^{22} 1.4661; $[\alpha]_D^{22}$ – 46.3° (c = 1.224, CHCl₃). The IR and nmr spectra were identical with those of the racemate. Higher work-up temp or distillation of the product caused racemization. (c) (R)-(+)-Isomer. In the same manner as above (S)-(+)-2 (1.12 g, $[\alpha]_{D}^{21.5}$ + 59.7° (MeOH)) yielded 800 mg (80%) of (R)-(+)-3a, $n_{D}^{21.5}$ 1.4671; $[\alpha]_{D}^{21.5}$ + 44.8° (c = 1.553, CHCl₃). This

showed the same spectral properties as those for (\pm) -3a.

3-Trimethylsilyloxymethyl-4-butanolide 3b

(a) (R)-(-)-Isomer. (Me₃Si)₂NH (1 ml) and Me₃SiCl (1 ml) were added to a stirred and ice-cooled soln of (S)-(-)-3a (990 mg, $[\alpha]_D^{22} - 46.3^\circ$ (CHCl₃)) in dry C₅H₅N (1 ml) and the mixture was kept at 0-5° for 2h. After dilution with C₆H₆-nhexane, the mixture was filtered through Celite to remove C₁H₅N · HCl. The filtrate was concentrated in vacuo. The residue was distilled to give 830 mg (52%) of (R)-3b, b.p. 140-144°/26 mm, n_{D}^{23} 1.4368; $[a]_{21}^{23}$ - 34.8° (c = 0.736, ether); ν_{max} 2950 (m), 2900 (m), 2860 (m), 1775 (s), 1470 (w), 1415 (m), 1370 (m), 1335 (w), 1250 (s), 1170 (s), 1090 (s), 1020 (m), 1000 (m), 870 (s), 840 (s), 745 (m), 680 (w) cm⁻¹; δ (CCl₄) 0.20 (9H, s), 2.25-3.05

(3H, m) 3.66 (2H, d, J-6 Hz), 4.10-4.70 (2H, m). (Found: C, 50.71; H, 8.35. Calc. for C₈H₁₆O₃Si: C, 51.03; H, 8.57%).

(b) (S)-(+)-Isomer. In the same manner as above (R)-(+)-3a (800 mg, $[\alpha]_D^{21.5} + 44.8^\circ$ (CHCl₃)) yielded 687 mg (52%) of (S)-3b, b.p. 140°/12 mm, $n_D^{21.5} + 1.4671; [\alpha]_D^{21.5} + 36.0^\circ$ (c = 0.86, ether). The IR and NMR spectra were identical with those of (R)-(-)-3b.

2-(6-Methylheptanoyl)-3-hydroxymethyl-4-butanolide (A-factor) 1a

(a) (3S)-(-)-Isomer (= natural A-factor). A soln of LiNPr¹₂ was prepared by the addition of n-BuLi (1.53 N in n-hexane, 7.3 ml) to a stirred and cooled soln of Pr¹₂ NH (1.1 g) in dry THF at -78° under Ar. To this was added with stirring and cooling at -78° a soln of (R)-3b (830 mg) in dry THF (5 ml) during 5 min. The soln was stirred at -78° for 10 min. Then 6-methylheptanovl chloride (0.8 g) in dry THF (2 ml) was added dropwise during 5 min at -78° . After stirring for 1 h at -78° , the mixture was poured into ice-AcOH-H₂O and extracted with ether. The ether soln was washed with H₂O, NaHCO₃ soln and brine, dried (MgSO₄) and concentrated in vacuo to give (3R)-1c as an oil with ν_{max} 1770 (s), 1720 (s), 865 (s), 840 (s) cm⁻¹. This was dissolved in 95% EtOH-H₂O (4:1, 15 ml) and the soln was stirred and heated under reflux for 10 min. After the removal of EtOH in vacuo, the product was extracted with ether. The ether soln was washed with H₂O and brine, dried (MgSO₄) and concentrated in vacuo. The residue was chromatographed over SiO₂ (Mallinckrodt Silicar CC-7, 6g) in n-hexane. Impurities were first eluted with n-hexane-ether (4:1) and then (3S)-1a was eluted with n-hexaneether (1:1) as 318 mg (30%) of a waxy solid, $[\alpha]_D^{23} - 13.1^\circ$ (c = 1.18, CHCl₃); v_{max} 3460 (m), 2960 (s), 2930 (s), 2860 (m), 1765 (s), 1720 (s), 1640 (w), 1470 (m), 1380 (m), 1365 (m), 1215 (m), 1170 (s), 1130 (m), 1095 (m), 1025 (s), 960 (w), 945 (w), 900 (w), 870 (w), 835 (w), 735 (w), 680 (w) cm⁻¹; δ (CDCl₃) 0.82 (6H, d, J = 6 Hz), 1.0-2.0 (9H, br), 2.5-3.3 (3H, m), 3.5-3.8 (2H, m), 3.9-4.2 (2H, m); ¹³C NMR (25 MHz, CDCl₃) 22.260, 23.167, 26.443, 27.466, 38.348, 39.225, 42.297, 54.758, 61.310, 69.120, 172.728 (C=O), 203.5 (C=O); MS: m/z 242.1604 (M⁺ = C₁₃H₂₂O₄, 3%), 211.1356 (C₁₂H₁₉O₃, 34%), 158.0531 (C₁H₁₀O₄, 30%), 153.0539 (C₈H₃O₃, 25%), 143.0461 (C₁₀H₇O, 20%), 141.0781 (C₁₁H₉, 28%), 127.1118 $\begin{array}{l} (C_8H_{15}O, 30\%), 127.0380 \ (C_6H_7O_3, 32\%), 116.0492 \ (C_5H_8O_3, 33\%), \\ 109.1039 \ (C_8H_{13}, 100\%), \ base \ peak), \ 102.0319 \ (C_4H_6O_3, 30\%), \\ 85.0278 \ (C_4H_5O_2, 77\%), \ 83.0853 \ (C_6H_{11}, 35\%), \ 82.0782 \ (C_6H_{10}, 46\%), \ 69.0709 \ (C_5H_9, 20\%), \ 67.0543 \ (C_3H_7, 28\%), \ 57.0716 \ (C_4H_9, 69\%), \ 55.0556 \ (C_4H_7, 33\%), \ 55.0191 \ (C_3H_3O, 29\%), \ 43.0573 \ (C_3H_7, 51\%), \ 41.0420 \ (C_3H_5, 40\%); \ CD \ (c = 0.062, \ MeOH): \ 283.5 \ nm \ (\Delta\epsilon + 0.699, \ peak), \ 221 \ nm \ (\Delta\epsilon + 0.420, \ peak). \end{array}$

(b) (3R)-(+)-Isomer. In the same manner as above (S)-3b (687 mg) yielded 256 mg (29%) of (3R)-(+)-1a, $[\alpha]_D^{22} + 12.7^\circ$ (CHCl₃); CD (c = 0.109, MeOH): 283.5 nm ($\Delta \epsilon - 0.593$, trough), 222 nm ($\Delta \epsilon - 0.402$, trough). This showed the same IR, ¹H NMR, ¹³C NMR and mass spectra as those of (3S)-(-)-1a.

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REFERENCES

- ¹A. S. Khokhlov, L. N. Anisova, I. I. Tovarova, E. M. Kleiner, I. V. Kovalenko, O. I. Krasilnikova, E. Ya, Kornitskaya and S. A. Pliner, Z. Allgem. Mikrobiol. 13, 647 (1973).
- ²E. M. Kleiner, S. A. Pliner, V. S. Soifer, V. V. Onoprienko, T. A. Balasheva, B. V. Rozynov and A. S. Khokhlov, *Bioorg. Khim.* 2, 1142 (1976).
- ³E. M. Kleiner, V. V. Onoprienko, S. A. Pliner, V. S. Soifer and A. S. Khokhlov, *Bioorg. Khim.* 3, 424 (1977).
- ⁴A. S. Khokhlov, Problems of studies of specific cell autoregulators. In Frontiers of Bioorganic Chemistry and Molecular Biology (Edited by S. N. Ananchenko), pp. 201–210. Pergamon Press, Oxford (1980).
- ⁵J. P. Jennings, W. Klyne and P. M. Scopes, J. Chem. Soc. 7211 (1965).
- ⁶Preliminary communication; K. Mori, *Tetrahedron Letters* 22, 3431 (1981).
- ⁷J. -F. Tocanne and C. Asselineau, Bull. Soc. Chim. France 3346 (1965).